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had mp 86–87°, $[\alpha]_D$ – 2° (c=1); (Found: C, 76.96; H, 10.06. $C_{32}H_{52}O_4$ requires: C, 76.75; H, 10.47%); IR ν_{max} cm⁻¹: 1720, 1710, 1250; PMR: δ 5.98 (t, 1H), 4.45 (m, 1H), 3.70, 2.10, 1.88 (three 3H, ss). Pure 2b, by hydrolysis with 10% hydromethanolic KOH and esterification with excess ethereal CH_2N_2 , was quantitatively converted into methyl 3-epiisomasticadienolate mp and mmp 140–141°, $[\alpha]_D$ – 16°, identical with an authentic specimen [5]. Further elution with petrol– C_6H_6 (1:1; 0.8 l.) gave the crude 1b (1400 mg) which after crystallization from MeOH had mp 129–130°, $[\alpha]_D$ – 26° (c=0.9); (Found: C, 76.12; H, 10.96. $C_{32}H_{52}O_4$ requires: C, 76.75; H, 10.47%); IR ν_{max} cm⁻¹: 1720, 1710, 1250; PMR: δ 5.88 (t, 1H), 5.20 (m, 1H), 4.45 (m, 1H), 3.70, 2.10, 1.88 (three 3H ss). Pure 1b, under the conditions reported above for 2b, afforded quantitatively methyl 3-epimasticadienolate (1d) mp and mmp 100–101°, $[\alpha]_D$ – 44°, identical with the natural 1d.

Masticadienonic aldehyde (1f). Evapn of the mother liquors (280 mg) of crystallization of oleanonic aldehyde afforded a semicrystalline solid which was recrystallized from MeOH to give crude 1f. Further purification of 1f was not possible since the substance was very prone to undergo visible alterations. IR $\nu_{\rm max}$ cm⁻¹: 1675; PMR: δ 9.45 (s, 1H, —CHO), 6.50 (m, 1H, C-24 vinyl H), 5.20 (m, 1H, C-7 vinyl H), 1.75 (s, 3H, Me on double bond). LAH reduction of crude 1f (100 mg) led to a crystalline product which after two crystallizations gave pure masticadienediol (1e) mp and mmp 186–187°, $[\alpha]_{\rm D}$ –50°, identical with an authentic specimen.

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REFERENCES

- Caputo, R., Mangoni, L., Monaco, P. and Palumbo, G. (1975) Phytochemistry 14, 809.
- Monaco, P., Caputo, R., Palumbo, G. and Mangoni, L. (1973) Phytochemistry 12, 946.
- Barton, D. H. R. and Seoane, E. (1956) J. Chem. Soc. 4150.
 Seoane, E. (1956) J. Chem. Soc. 4158.
- 5. Caputo, R. and Mangoni, L. (1970) Gazz. Chim. Ital. 100, 317.
- Monaco, P., Caputo, R., Palumbo, G. and Mangoni, L. (1973) Phytochemistry 12, 2534.
- 7. Cheung, H. T. and Feng, M. C. (1968) J. Chem. Soc. 4150.
- Arigoni, D., Jeger, O. and Ruzicka, R. (1955) Helv. Chim. Acta 38, 222.
- Crabbe, P., Ourisson, G. and Takahashi, T. (1958) Tetrahedron 3, 279.
- 10. Mills, J. S. (1956) J. Chem. Soc. 2196.
- 11. Monaco, P., Caputo, R., Palumbo, G. and Mangoni, L. (1973) Phytochemistry 12, 939.
- Simonsen, G. and Ross, L. (1951) The Terpenes Vol. 4. Cambridge University Press, London.
- Monaco, P., Caputo, R., Palumbo, G. and Mangoni, L. (1974) Phytochemistry 13, 1992.
- Shamma, M. and Rosenstock, P. D. (1959) J. Org. Chem. 24, 726.

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ON THE OCCURRENCE OF FILICAN-3-ONE IN STRYCHNOS DOLICHOTHYRSA

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Key Word Index-Strychnos; Loganiaceae; filican-3-one; triterpene.

Plant material of Strychnos dolichothyrsa Gilg ex Onochie et Hepper (Loganiaceae) was collected by Dr A. J. M. Leeuwenberg and Prof. F. Sandberg in the vicinity of Kribi, Cameroun (collection number Lg 7870) [1, 2]. A voucher specimen is kept in the herbarium at Wageningen, The Netherlands. The stem bark of this species was investigated for its muscle relaxant alkaloids [3, 4]. Prior to the alkaloid extraction the ground stem bark was extracted with hexane. Upon evaporation of the hexane, a crystalline compound was formed. The isolated compound proved to be a saturated pentacyclic triterpene, filican-3-one (1) (D: A. Friedo-B': A'-neogammaceran-3-one), its structure and its stereochemistry having been established by X-ray crystallography [5]. Filican-3-one has been described previously in connection with the structure elucidation of triterpenes with the filicane skeleton [6, 7]. However, to our knowledge its natural occurrence has not been reported before.

Filican-3-one (recrystallized $\times 2$ ether) mp 248-249° (some sublimation was observed). $[\alpha]_D^{25^\circ}$ -25.4, no

Filican-3-one (1)

depression of the mmp with an authentic sample of filican-3-one (kindly provided by Prof. A. Marsili, Pisa, Italy). High resolution mass spectrometry (direct inlet, 240°, 70 eV) m/e M⁺ ($C_{30}H_{50}O$) 426 (32), 411 (36), 383 (4) M⁺-isopropyl, 341 (44) $C_{24}H_{37}O^+$, 274 (18) $C_{19}H_{30}O^+$, 273 (20) $C_{20}H_{33}^+$ and $C_{19}H_{29}O^+$ (1:3), 259 (20) $C_{18}H_{27}O^+$, 257 (20) $C_{18}H_{25}O^+$, 234 (56) $C_{16}H_{26}O^+$ and $C_{17}H_{19}^+$ (1:10), 233 (32) $C_{16}H_{25}O^+$ and $C_{17}H_{29}^+$ (1:1), 231 (16) $C_{16}H_{23}O$, 219 (20) $C_{15}H_{23}O$, 206 (36)

 $C_{14}H_{22}O^+$ and $C_{15}H_{26}^+$ (1:5), 205 (40) $C_{14}H_{21}O$, † 191 (76) $C_{13}H_{19}O^+$ and $C_{14}H_{21}^+$ (1:10), 179 (44) $C_{12}H_{19}O^+$ and $C_{13}H_{23}^+$ (1:2), 178 (28), 177 (24), 149 (34), 137 (48) $C_{9}H_{13}O^+$ and $C_{10}H_{17}^+$ (1:8), 135 (40), 123 (60) $C_{8}H_{11}O^+$ and $C_{9}H_{15}^+$ (1:7), 121 (74), 109 (100) and 107 (48).

The peaks at m/e 302, 246 and 218, present in the MS of friedelin, (2) are not observed in that of filican-3-one. Other fragments as 341 (loss of E-ring) and 205 contain oxygen, unlike the fragments m/e 341 and 205 in friedelin. The fragments 341, 234 and 191 are more abundant in the mass spectrum of filican-3-one than in the spectrum of friedelin; the opposite is true for the 232 and 125 fragments [8-10]. These differences distinguish filican-3-one clearly from friedelin.

IR spectrum v_{max} (KBr-disc): 3400, 2900, 1705, 1455, 1380, 1260, 1200, 1070, 1005 and 800 cm⁻¹. NMR spectrum (100 MHz in CDCl₃): the most upfield signal is at δ 0.71 ppm, like in friedelin it is assigned to 5-Me [11, 12]. Based on the integration, stereochemical considerations and the shifts observed for the Me groups upon increasing concentration of the shift reagent Eu (FOD)₃, the following assignments are made for the methyl signals: δ in ppm 0.71 (5-Me), 0.80 (17-Me), 0.83 (d, J = 6.5 Hz, 22-Me), 0.875 (d, J = 6.5 Hz, 4-Me),

0.89 (d, J=6.5 Hz, 22-Me), 0.915 (6 H, 13-Me and 9-Me) and 0.99 (14-Me). In deuterobenzene the NMR spectrum showed the following shift for the methyl groups: δ in ppm, 0.68 (5-Me), 0.75 (17-Me), 0.835 (9-Me), 0.87 (13-Me), 0.901 (d, J=6.5 Hz, 22-Me), 0.96 (d, J=6.5 Hz, 22-Me), 0.975 (d, J=6.5 Hz, 4-Me), and 1.01 (14-Me). This means that the 4-Me signal is shifted 0.10 ppm to the lower field, which is in accordance with the shift observed for friedelin, under the same circumstances [12]. This confirms the β -configuration of the 4-Me (= equatorial).

REFERENCES

- Leeuwenberg, A. J. M. (1969) Mededel. Landbouwhogeschool Wageningen 69, 1.
- Sandberg, F., Verpoorte, R. and Cronlund, A. (1971) Acta Pharm. Suecica 8, 341.
- Verpoorte, R. and Baerheim Svendsen, A. (1976) Lloydia 39, 357.
- 4. Verpoorte, R. (1976) Thesis, State University Leiden, The Netherlands.
- 5. Pedersen, B. F. and Verpoorte, R. to be published.
- 6. Berti, G., Bottari, F and Marsili, A. (1969) Tetrahedron 25,
- Ageta, H., Iwata, K. and Natori, S. (1964) Tetrahedron Letters 3413.
- 8. Courtney, J. L. and Shannon, J. S. (1963) Tetrahedron Letters 13.
- Courtney, J. L. and Shannon, J. S. (1963) Tetrahedron Letters 173.
- Budzikiewicz, H., Wilson, J. M. and Djerassi, C. (1963) J. Am. Chem. Soc. 85, 3688.
- 11. Ageta, H. and Iwata, K. (1966) Tetrahedron Letters 6069.
- Aoyagi, R., Yamada, S., Tsuyuki, T. and Takahashi, T. (1973) Bull. Chem. Soc. Japan 46, 959.

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CAROTENOID SYNTHESIS IN TURKISH LEMONS AND ORANGES AS INFLUENCED BY TRIETHYLAMINE DERIVATIVES

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Key Word Index—Citrus; Rutaceae; lemons; oranges; carotenoids; triethylamines; lycopene.

Abstract—Two triethylamines, 2-(4-chloro-phenylthio)-triethylamine (CPTA) and 4-chloro[β -(diethylamino)-ethyl]-benzoate (CDEB), were used to investigate C_{30} carotenoid biosynthesis in Turkish oranges and lemons. It was not possible to determine the pathway, since not all the intermediates from phytoene to the C_{30} compounds were observed. Lycopene and ζ -carotene, absent in controls, were identified in large amounts in certain CPTA- and CDEB-treated lemons and oranges. Furthermore there were varietal differences, since CDEB-treated Turkish oranges did not produce the increases in α - and β -carotene earlier observed in Californian Citrus fruits.

INTRODUCTION

Yokoyama et al. [1-3] have shown that CPTA affected carotenoid biosynthesis by increasing lycopene produc-

tion whilst CDEB increased β -carotene over a certain period [4]. Since it is known that varietal differences played an important role in response to these compounds